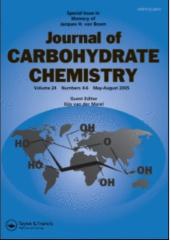
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SYNTHESIS OF  $\beta$ -d-Glcp-(1 $\rightarrow$ 2)-[ $\beta$ -d-Ribf-(1 $\rightarrow$ 3)-] $\alpha$ -l-Rhap-(1 $\rightarrow$ 3)- $\alpha$ -l-Rhap-(1 $\rightarrow$ 2)- $\alpha$ -l-Rhap, THE REPEATING UNIT OF THE LIPOPOLYSACCHARIDE OF Acetobacter diazotrophicus PAL 5

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# SYNTHESIS OF $\beta$ -D-Glcp- $(1 \rightarrow 2)$ - $[\beta$ -D-Ribf- $(1 \rightarrow 3)$ - $]\alpha$ -L-Rhap- $(1 \rightarrow 3)$ - $\alpha$ -L-Rhap- $(1 \rightarrow 2)$ - $\alpha$ -L-Rhap, THE REPEATING UNIT OF THE LIPOPOLYSACCHARIDE OF ACETOBACTER DIAZOTROPHICUS PAL 5

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## ABSTRACT

A pentasaccharide, the major repeating unit of the lipopolysaccharide (LPS) of the nitrogen fixing bacterium Acetobacter diazotrophicus PAL 5 was efficiently synthesized as its allyl glycoside using a regio- and stereoselective strategy. The key acceptor, allyl 3-O-acetyl-4-O-benzoyl-α-Lrhamnopyranoside (3), was prepared by selective 3-O-acetylation of allyl 4-O-benzoyl-α-L-rhamnopyranoside. Condensation of 3 with 2,3,4,6-tetra-Obenzoyl-α-D-glucopyranosyl trichloroacetimidate furnished the disaccharide 5. Deallylation and subsequent trichloroacetimidation of 5 afforded 2,3,4,6tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-benzoyl- $\alpha$ -Lrhamnopyranosyl trichloroacetimidate (10). Selective 3-O-glycosylation of allyl  $\alpha$ -L-rhamnopyranoside (1) with 10 followed by benzoylation gave trisaccharide (12), which could be conveniently converted to a donor (14). Condensation of 14 with ally 3,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranoside (15) gave tetrasaccharide 16. Selective deacetylation of 16 gave the acceptor 17 which was ribosylated to furnish the protected pentasaccharide, and finally deprotection led to the title compound.

Key Words: Regio- and stereoselective synthesis; Rhamnan; Trichloroacetimidate

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## INTRODUCTION

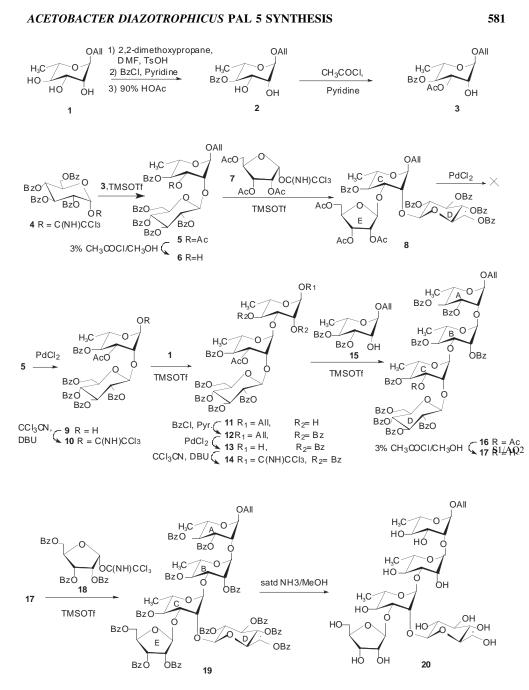
The pentasaccharide,  $\beta$ -D-Glc*p*- $(1 \rightarrow 2)$ - $[\beta$ -D-Rib*f*- $(1 \rightarrow 3)$ - $]\alpha$ -L-Rha*p*- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha*p*- $(1 \rightarrow 2)$ - $\alpha$ -L-Rha*p*, is the major repeating unit of the lipopolysaccharide (LPS) of the nitrogen fixing bacterium *Acetobacter diazotrophicus* PAL 5 occurring in sugar cane in Brazil and Australia.<sup>[11]</sup> Field experiments suggest that up to 80% of the N incorporated into Brazilian sugar cane may be obtained from biological nitrogen fixation (BNF). The involvement of the carbohydrate-rich molecules in establishing the interaction between the BNF bacterium and the host has been reported.<sup>[2,3]</sup> These facts are of interest from the viewpoints of the biological roles of carbohydrates.

Rhamnans with a long backbone consisting of  $\alpha$ -(1 $\rightarrow$ 2) and  $\alpha$ -(1 $\rightarrow$ 3) linked L-rhamnose units to which are attached various kinds of side chains are widely distributed in nature. These target compounds are structurally very similar, but the synthetic approaches used are quite different. A stepwise synthesis of the hexasaccahride with rhamnotetraose as the backbone and two glucosamine units as the side chains has been reported.<sup>[4]</sup> Our previous work described highly regio- and stereoselective syntheses of oligosaccharides via orthoester formation-rearrangement strategy using glycosyl trichloroacetimidates as the donors and lightly protected sugars as the acceptors were achieved in a one-pot manner.<sup>[5–7]</sup> As part of our ongoing research project on the synthesis of rhamnans, we present herein the synthesis of the well-defined ribosylated glucorhamnan pentasaccharide.

#### **RESULTS AND DISCUSSION**

As outlined in Scheme 1, allyl  $\alpha$ -L-rhamnopyranoside was converted to allyl 4-Obenzoyl- $\alpha$ -L-rhamnopyranoside (2) through 2,3-O-isopropylidenation with 2,2-dimethoxypropane in DMF in the presence of catalytic TsOH, 4-O-benzoylation with benzoyl chloride in pyridine, and deisopropylidenation with 90% acetic acid. These three steps were performed continuously without separation, giving 85.3% overall yield. Subsequent acetylation of 2 with 1.05 equiv of acetyl chloride in pyridinedichloromethane selectively gave allyl 3-O-acetyl-4-O-benzoyl-\alpha-L-rhamnopyranoside (3) as the only product in 91.5% yield. Its structure was confirmed by its <sup>1</sup>H NMR spectrum which showed a characteristic upfield signal at  $\delta$  4.11 ppm (dd, J<sub>1,2</sub>=1.6 Hz,  $J_{2,3}=3.2$  Hz) for H-2. Coupling of the glucose donor 4 with the acceptor 3 in the presence of catalytic TMSOTf furnished the  $(1 \rightarrow 2)$ -linked disaccharide 5. The 3-Oacetyl group of the disaccharide 5 was successfully removed without affecting any of the benzoyl groups in CH<sub>3</sub>COCl-methanol<sup>[8]</sup> (3%) to give the desired acceptor 6 in 84.5% yield. Thus coupling of 6 with 2,3,4-tri-O-acetyl-α-D-ribofuranosyl trichloroacetimidate<sup>[9]</sup> (7) gave trisaccharide 8 in satisfactory yield (79.5%). However, perhaps because of the presence of peracetylated ribofuranosyl group, an attempt to deallylate 8 with PdCl<sub>2</sub> was not successful and gave a very complex product. Later on, we tried to first construct the glucorhamnose tetrasaccharide acceptor 17, and then successively made the target pentasaccharide. Thus, deallylation of 5 with PdCl<sub>2</sub>, followed by trichloroacetimidation<sup>[10]</sup> with CCl<sub>3</sub>CN in the presence of DBU or K<sub>2</sub>CO<sub>3</sub> gave the disaccharide donor 10. Coupling of the donor 10 with the acceptor allyl  $\alpha$ -L-rhamnopyranoside (1) in the presence of catalytic TMSOTf selectively gave the  $(1 \rightarrow 3)$ linked trisaccharide 11. Benzoylation of 11 gave allyl 2,3,4,6-tetra-O-benzoyl- $\beta$ -D-

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Scheme 1. Synthesis of the target pentasaccharide.

glucopyranosyl- $(1 \rightarrow 2)$ -3-*O*-acetyl-4-*O*-benzoy- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 3)$ -2,4-di-*O*-benzoy- $\alpha$ -L-rhamnopyranoside (**12**, 68.1% for 2 steps). The <sup>1</sup>H NMR spectrum of **12** showed characteristic upfield signals at  $\delta$  4.45 ppm (dd, J<sub>2,3</sub>=3.0 Hz, J<sub>3,4</sub>=9.8 Hz) for H-3 and  $\delta$  3.75 ppm (dd, J<sub>1,2</sub>=1.3 Hz, J<sub>2,3</sub>=3.2 Hz) for H-2 respectively. It was noted that the temperature during addition of TMSOTf had to be maintained below  $-20^{\circ}$ C

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to ensure formation of the orthoester intermediate, otherwise, for example at room temperature, the regioselectivity was poor. Condensation of the rhamnose donor 14, readily prepared from 12 through deallylation and trichloroacetimidation, with the acceptor 15 furnished the tetrasaccharide 16. Subsequent selective deacetylation of 16 with CH<sub>3</sub>COCl-methanol (3%) afforded the tetrasaccharide acceptor, allyl 2,3,4,6-tetra-*O*-benzoyl- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 2)-4-*O*-benzoyl- $\alpha$ -L-rhamnopyranosyl-(1 $\rightarrow$ 2)-3,4-di-*O*-benzoyl- $\alpha$ -L-rhamnopyranosyl (1 $\rightarrow$ 2)-3,4-di-*O*-benzoyl- $\alpha$ -L-rhamnopyranosyl trichloroacetimidate<sup>[9]</sup> (18) proceeded smoothly in dichloromethane in the presence of TMSOTf, giving the pentasaccharide (19). The structure of 19 was specified by its <sup>1</sup>H, <sup>13</sup>C NMR, and <sup>1</sup>H-<sup>1</sup>H COSY NMR spectra. Deacylation of 19 in ammonium-saturated methanol gave the target pentasaccharide 20. The bioassay of 20 is in progress.

In summary, a very concise and efficient synthesis of allyl  $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -[ $\beta$ -D-ribofuranosyl- $(1 \rightarrow 3)$ -] $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)$ - $\alpha$ -L-rhamnopyranoside was achieved through a regio- and stereoselective process. In terms of the simplicity and efficiency, this method can be used for construction of higher oligosaccharides with similar structures.

### **EXPERIMENTAL**

General methods. Melting points were determined with a 'Mel-Temp' apparatus. Optical rotations were determined with a Perkin-Elmer model 241-MC automatic polarimeter for solutions in a 1-dm, jacketed cell. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded with Varian XL-400 and Varian XL-200 spectrometers, for solutions in CDCl<sub>3</sub> with tetramethylsilane (Me<sub>4</sub>Si) as the internal standard or in  $D_2O$  with acetone as the internal standard. Chemical shifts are expressed in ppm downfield from the internal Me<sub>4</sub>Si absorption. Mass spectra were recorded with a VG PLATFORM mass spectrometer using the ESI mode. Thin-layer chromatography (TLC) was performed on silica gel HF, detection being affected by charring with 30% (v/v) sulfuric acid in methanol or sometimes by UV detection. Column chromatography was conducted by elution of a column ( $16 \times 240$ ,  $18 \times 300$ ,  $35 \times 400$  mm) of silica gel (100-200 mesh) with EtOAc-petroleum ether ( $60-90^{\circ}$ C) as the eluent. Analytical LC was performed with a Gilson HPLC consisting of a pump (model 306), stainless steel packed with silica gel (Spherisorb SiO<sub>2</sub>,  $10 \times 300$  mm or  $4.6 \times 250$  mm), differential refractometer (132-RI Detector), UV/VIS detector (model 118), and EtOAc/petroleum ether (bp 60- $90^{\circ}$ C) was used as the eluent at a flow rate of 1–4 mL/min. Solutions were concentrated at a temperature  $<60^{\circ}$ C under diminished pressure.

Allyl 4-O-benzoyl- $\alpha$ -L-rhamnopyranoside (2). To a solution of allyl  $\alpha$ -L-rhamnopyranoside (1) (2.04 g, 10 mmol) in DMF (10 mL) containing *p*-toluenesulfonic acid monohydrate (38 mg, 0.2 mmol) was added 2,2-dimethoxypropane (2.5 mL, 20 mmol) and the mixture was stirred for 12 h, at the end of which time TLC (3/1 petroleum ether/ethyl acetate) indicated that the reaction was complete. Then the reaction mixture was added dropwise to a solution of pyridine (20 mL) containing benzoyl chloride (4.7 mL, 40 mmol). After stirring for 24 h at room temperature, the mixture was diluted

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with dichloromethane, washed with 1 N hydrochloric acid, water, and satd aq solution of sodium bicarbonate subsequently. The organic layer was combined, dried, and concentrated to a residue. The residue was dissolved in 90% acetic acid and refluxed for 1 h. The solution was concentrated, and purification of the residue by flash column chromatography on a silica gel column (1:1 petroleum ether–EtOAc) gave compound **2** (2.63 g, 85.4%) as a syrup;  $[\alpha]_D - 71.3^\circ$  (*c* 1.3, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  8.06–7.42 (m, 5 H, Bz-H), 5.93 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.32–5.21 (m, 2 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.07 (dd, 1 H, J<sub>3,4</sub>=J<sub>4,5</sub>=9.9 Hz, H-4), 4.92 (d, 1 H, J<sub>1,2</sub>=0.8 Hz, H-1), 4.24–3.98 (m, 5 H), 3.07–2.90 (bs, 2 H, 2 OH), 1.24 (d, J<sub>5,6</sub>=6.4 Hz, 3 H, H-6).

Anal. Calcd for C<sub>16</sub>H<sub>.20</sub>O<sub>6</sub>: C, 62.32; H, 6.54. Found: C, 64.49; H, 6.76.

Allyl 3-*O*-Acetyl-4-*O*-benzoyl-α-L-rhamnopyranoside (3). A solution of acetyl chloride (0.8 mL, 11 mmol) in anhyd dichloromethane (10 mL) was added dropwise to a solution of allyl 4-*O*-benzoyl-α-L-rhamnopyranoside (2, 3.08 g, 10 mmol) in anhyd dichloromethane (100 mL) containing 5 mL pyridine at 0°C within 10 min, and the mixture was stirred at room temperature for 2 h, at the end of which time TLC (3/1 petroleum ether/ethyl acetate) indicated that the reaction was complete. The mixture was washed with 1 N hydrochloric acid, water, and satd aq solution of sodium bicarbonate subsequently. The organic layer was combined, dried, and concentrated to a residue. Purification of the residue on a silica gel column with 3:1 petroleum ether–ethyl acetate as the eluent gave **3** (3.20 g, 91.5%) as a syrup;  $[\alpha]_D - 53.2^\circ$  (*c* 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 8.06–7.42 (m, 5 H, Bz-H), 5.93 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.48 (dd, 1 H, J<sub>2,3</sub>=3.2 Hz, J<sub>3,4</sub>=9.8 Hz, H-3), 5.37 (dd, 1 H, J<sub>3,4</sub>=J<sub>4,5</sub>=9.8 Hz, H-4), 5.36–5.23 (m, 2 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 4.90 (d, 1 H, J<sub>1,2</sub>=1.6 Hz, H-1), 4.24 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 4.11 (dd, 1 H, J<sub>1,2</sub>=1.6 Hz, J<sub>2,3</sub>=3.2 Hz, H-2), 4.07–4.01 (m, 2 H, H-5, OCH<sub>2</sub>CHCH<sub>2</sub>), 1.99 (s, 3 H, CH<sub>3</sub>CO), 1.25 (d, J<sub>5,6</sub>=6.3 Hz, 3 H, H-6).

Anal. Calcd for C<sub>18</sub>H<sub>.22</sub>O<sub>7</sub>: C, 61.70; H, 6.33. Found: C, 61.86; H, 6.30.

Allyl 2,3,4,6-Tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-ben**zoyl-\alpha-L-rhamnopyranoside** (5). 2,3,4,6-Tetra-O-benzoyl- $\alpha$ -D-glucopyranosyl trichloroacetimidate (4) (3.70 g, 5 mmol) and allyl 3-O-acetyl-4-O-benzoyl- $\alpha$ -L-rhamnopyranoside (3) (1.75 g, 5 mmol) were dried together under high vacuum for 2 h, then dissolved in anhyd CH<sub>2</sub>Cl<sub>2</sub> (20 mL). TMSOTf (45 µL, 0.05 equiv) was added dropwise at  $-10^{\circ}$ C with N<sub>2</sub> protection. The reaction mixture was stirred for 3 h, during which time the temperature was gradually warmed to ambient temperature. Then the mixture was neutralized with triethylamine and concentrated to dryness. Purification by column chromatography (3:1 petroleum ether-EtOAc) gave 5 (3.92 g, 84.5%) as a foamy solid;  $[\alpha]_D = 5.8^{\circ}$  (c 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.15–7.43 (m, 25 H, 5 PhH), 5.96 (dd, 1 H, J<sub>2.3</sub>=J<sub>3.4</sub>=9.8 Hz, H-3'), 5.80 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.66 (dd, 1 H,  $J_{3,4} = J_{4,5} = 9.8$  Hz, H-4'), 5.61 (dd, 1 H,  $J_{1,2} = 7.8$  Hz,  $J_{2,3} = 9.8$  Hz, H-2'), 5.38 (dd, 1 H, J<sub>2.3</sub>=3.2 Hz, J<sub>3.4</sub>=9.7 Hz, H-3), 5.39-5.21 (m, 3 H, H-4, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.04 (d, 1 H,  $J_{1,2}=1.4$  Hz, H-1), 4.88 (d, 1 H,  $J_{1,2}=7.8$  Hz, H-1'), 4.60-4.46 (m, 2 H, H-6'), 4.15-3.88 (m, 5 H, H-2, H-5, H-5', OCH<sub>2</sub>CHCH<sub>2</sub>), 1.26 (s, 3 H, COCH<sub>3</sub>), 1.22 (d, 3 H, J<sub>5.6</sub>=6.4 Hz, H-6); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 170.2 (COCH<sub>3</sub>), 165.9, 165.8, 165.1, 164.9, 164.8 (5 C, 5 COPh), 117.4 (OCH2CHCH2), 102.8, 98.1 (2 C, C-1, 1'), 78.3, 72.4, 72.2, 71.9, 71.8, 70.4, 69.5, 68.2, 66.7, 63.0, 19.7, 16.5.

Anal. Calcd for C<sub>52</sub>H<sub>48</sub>O<sub>16</sub>: C, 67.23; H, 5.21. Found: C, 67.09; H, 5.40.

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Allyl 2,3,4,6-Tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -4-O-benzoyl- $\alpha$ -Lrhamnopyranoside (6). To a solution of 5 (1.86 g, 2 mmol) in anhyd MeOH (50 mL) was added CH<sub>3</sub>COCl (1.5 mL) at 0°C. The solution was stoppered in a flask and stirred at room temperature until TLC (3:1 petroleum ether-EtOAc) showed that the starting material disappeared. The solution was neutralized with Et<sub>3</sub>N, then concentrated to dryness. The residue was passed through a short silica gel column to give  $\mathbf{6}$ (1.69 g, 94.9%) as a white solid;  $[\alpha]_{\rm D}$  + 18.5° (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.03-7.27 (m, 25 H, 5 PhH), 5.95 (dd, 1 H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.6 Hz, H-3'), 5.80 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.70–5.63 (m, 2 H, H-2', H-4'), 5.25–5.13 (m, 3 H, H-1', OCH<sub>2</sub>CHCH<sub>2</sub>), 5.04 (d, 1 H, J<sub>1,2</sub>=0.8 Hz, H-1), 4.90 (dd, 1 H, J<sub>3,4</sub>=J<sub>4,5</sub>=9.8 Hz, H-4), 4.67-4.50 (m, 2 H, H-6'), 4.20 (m, 1 H, H-5), 4.10 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 4.03 (dd, 1 H, J<sub>1,2</sub>=0.8 Hz, J<sub>2,3</sub>=3.1 Hz, H-2), 4.00 (dd, 1 H, J<sub>2,3</sub>=3.1 Hz, J<sub>3,4</sub>=9.8 Hz, H-3), 3.92-3.84 (m, 2 H, H-5', OCH<sub>2</sub>CHCH<sub>2</sub>), 1.21 (d, 3 H, J<sub>5.6</sub>=6.3 Hz, H-6); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 166.5, 166.0, 165.7, 165.2, 165.1 (5 C, 5 COPh), 117.1 (OCH<sub>2</sub>CHCH<sub>2</sub>), 102.9, 98.1 (2 C, C-1, 1'), 80.3, 75.4, 72.6, 72.3, 72.1, 70.0, 69.5, 68.1, 66.0, 63.1, 17.4.

Anal. Calcd for C<sub>50</sub>H<sub>46</sub>O<sub>15</sub>: C, 67.71; H, 5.23. Found: C, 67.53; H, 5.42.

Allyl 2,3,4-Tri-O-acetyl- $\beta$ -D-ribofuranosyl- $(1 \rightarrow 3)$ -[2,3,4,6-tetra-O-benzoyl- $\beta$ -**D-glucopyranosyl-(1\rightarrow2)]-\alpha-L-rhamnopyranoside (8). 2,3,4-Tri-O-acetyl-\alpha-D-ribo**furanosyl trichloroacetimidate (7, 421 mg, 1.0 mmol) and allyl 2,3,4,6-tetra-O-benzoyl-β-D-glucopyranosyl- $(1 \rightarrow 2)$ -4-O-benzoyl-α-L-rhamnopyranoside (6, 886 mg, 1.0 mmol) were dried together under high vacuum for 2 h, then dissolved in anhyd  $CH_2Cl_2$ (10 mL). TMSOTf (18  $\mu$ L, 0.10 mmol) was added dropwise at  $-20^{\circ}$ C with N<sub>2</sub> protection. The reaction mixture was stirred for 3 h, during which time the temperature was gradually warmed to ambient temperature. Then the mixture was neutralized with triethylamine and concentrated to dryness. Purification of the residue by column chromatography (1:1 petroleum ether-EtOAc) gave 8 (910 mg, 79.5%) as a foamy solid.  $[\alpha]_{D} + 9.3^{\circ}$  (c 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.17–7.28 (m, 25 H, 5 PhH), 5.95 (dd, 1 H, J<sub>3,4</sub>=J<sub>4,5</sub>=9.7 Hz, H-4<sub>C</sub>), 5.83 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.69 (dd, 1 H,  $J_{3,4} = J_{4,5} = 9.7$  Hz, H-4<sub>D</sub>), 5.60 (dd, 1 H,  $J_{2,3} = 9.6$  Hz,  $J_{3,4} = 9.7$  Hz, H-3<sub>D</sub>), 5.36 (d, 1 H,  $J_{2,3}=3.7$  Hz, H-2<sub>E</sub>), 5.26 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.21 (d, 1 H,  $J_{1,2}=7.7$  Hz, H-1<sub>D</sub>), 5.16–5.08 (m, 3 H, H-1<sub>C</sub>, H-2<sub>D</sub>, OCH<sub>2</sub>CHCH<sub>2</sub>), 4.96–4.92 (m, 2 H, H-1<sub>E</sub>,  $H-3_E$ ), 4.66–4.67 (m, 2 H,  $H-6_D$ ), 4.30–3.70 (m, 8 H), 2.09 (s, 3 H,  $COCH_3$ ), 2.04 (s, 3 H, COCH<sub>3</sub>), 1.40 (s, 3 H, COCH<sub>3</sub>), 1.08 (d, 3 H,  $J_{5,6}$ =6.3 Hz, H-6<sub>C</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 170.4, 169.8, 169.6 (3 C, 3 COCH<sub>3</sub>), 166.0, 165.8, 165.3, 165.3, 165.1, 165.0 (5 C, 5 COPh), 116.9 (OCH<sub>2</sub>CHCH<sub>2</sub>), 102.0, 98.4, 97.6 (3 C, C-1<sub>C</sub>, 1<sub>D</sub>, 1<sub>E</sub>), 78.3, 73.9, 72.6, 72.1, 72.0, 71.9, 70.0, 69.8, 68.0, 67.2, 66.8, 65.0, 63.1, 62.1, 20.9, 20.8, 20.6, 17.5.

Anal. Calcd for C<sub>61</sub>H<sub>60</sub>O<sub>22</sub>: C, 63.98; H, 5.28. Found: C, 63.95; H, 5.51.

2,3,4,6-Tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-benzoyl- $\alpha$ -L-rhamnopyranosyl trichloroacetimidate (10). To a solution of allyl 2,3,4,6-tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-benzoyl- $\alpha$ -L-rhamnopyranoside (5, 928 mg, 1 mmol) in 90% acetic acid (10 mL) containing sodium acetate (293 mg, 3 mmol) was added PdCl<sub>2</sub> (89 mg, 0.5 mmol), and the mixture was stirred for 12 h,

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at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the reaction was complete. The mixture was diluted with dichloromethane (30 mL), washed with water and satd aq sodium bicarbonate. The organic layer was concentrated, and the residue was passed through a short silica gel column with 2:1 petroleum ether-EtOAc as the eluent to give crude 2,3,4,6-tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-benzoyl- $\alpha$ ,  $\beta$ -L-rhamnopyranose (9, 844 mg, 95.0%). Compound 9 was dissolved in dichloromethane (10 mL), and CCl\_3CN (0.2 mL, 2 mmol) and DBU (27  $\mu L,~0.18$ mmol) were added. The reaction mixture was stirred for 2 h, at the end of which time TLC (3:1 petroleum ether-ethyl acetate) indicated that the reaction was complete. Concentration of the reaction mixture followed by purification on a silica gel column with 3:1 petroleum ether-EtOAc as the eluent, furnished the disaccharide donor 10 (876 mg, 89.4%) as a foamy solid;  $[\alpha]_D - 11.6^\circ$  (c 1.1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.42 (s, 1 H, CNHCCl<sub>3</sub>), 8.17-7.28 (m, 25 H, 5 PhH), 6.52 (d, 1 H, J<sub>1,2</sub>=1.6 Hz, H-1), 5.99 (dd, 1 H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.7 Hz, H-3'), 5.70–5.59 (m, 2 H, H-4', H-2'), 5.37 (dd, 1 H,  $J_{3,4}=J_{4,5}=9.8$  Hz, H-4), 4.93 (d, 1 H,  $J_{1,2}=7.7$  Hz, H-1'), 4.63–4.53 (m, 2 H, H-3, H-6'), 4.32 (dd, 1 H,  $J_{1,2}=1.6$  Hz,  $J_{2,3}=3.1$  Hz, H-2), 4.24-4.12 (m, 2 H, H-5, H-6'), 1.30 (d, 3 H, J<sub>5.6</sub>=6.4 Hz, H-6), 1.22 (s, 3 H, COCH<sub>3</sub>).

Anal. Calcd for C<sub>51</sub>H<sub>44</sub>Cl<sub>3</sub>NO<sub>16</sub>: C, 59.28; H, 4.29. Found: C, 59.50; H, 4.44.

Allyl 2,3,4,6-Tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-benzoyl-α-L-rhamnopyranosyl- $(1 \rightarrow 3)$ -2,4-di-*O*-benzoyl-α-L-rhamnopyranoside (12). 2,3,4,6-Tetra-*O*-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-*O*-acetyl-4-*O*-benzoyl- $\alpha$ -L-rhamnopyranosyl trichloroacetimidate (10, 1.03 g, 1 mmol) and allyl  $\alpha$ -L-rhamnopyranoside (1, 204 mg, 1 mmol) were dried together under high vacuum for 2 h, then dissolved in anhyd CH<sub>2</sub>Cl<sub>2</sub> (40 mL). TMSOTf (18  $\mu$ L, 0.1 mmoL) was added dropwise at  $-25^{\circ}$ C with  $N_2$  protection. The reaction mixture was stirred for 3 h, during which time the temperature was gradually warmed to ambient temperature. Then the mixture was neutralized with triethylamine, and concentrated to dryness under reduced pressure to afford the crude allyl 2,3,4,6-tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-*O*-benzoyl- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 3)$ - $\alpha$ -L-rhamnopyranoside (11). To the solution of crude 11 in pyridine (20 mL) was added benzoyl chloride (3.5 mL, 30 mmol) dropwise, and the mixture was stirred overnight at room temperature. TLC (3:1 petroleum ether-EtOAc) indicated that the reaction was complete. Ice water was added, and the mixture was diluted with dichloromethane, washed with 1 N hydrochloric acid, water, and satd aq sodium bicarbonate. The organic layer was combined, dried, and concentrated. Purification of the crude product by column chromatography (3:1 petroleum ether-EtOAc) gave 12 (810 mg, 68.1% for 2 steps) as a syrup:  $[\alpha]_{\rm D} + 42.6^{\circ}$  (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.15–7.27 (m, 35 H, 7 PhH), 5.95 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.71 (dd, 1 H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.6 Hz, H-3"), 5.56-5.46 (m, 2 H, H-4", H-2"), 5.44 (dd, 1 H,  $J_{1,2}=1.5$  Hz,  $J_{2,3}=3.1$  Hz, H-2), 5.42–5.25 (m, 3 H, H-4, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.13 (dd, 1 H, J<sub>2.3</sub>=3.2 Hz, J<sub>3.4</sub>=9.6 Hz, H-3'), 5.11 (d, 1 H, J<sub>1.2</sub>=1.5 Hz, H-1/H-1'), 5.06 (d, 1 H,  $J_{1,2}$ =1.0 Hz, H-1/H-1'), 5.05 (dd, 1 H,  $J_{3,4}$ = $J_{4,5}$ =9.6 Hz, H-4'), 4.48 (d, 1 H, J<sub>1,2</sub>=7.7 Hz, H-1"), 4.45 (dd, 1 H, J<sub>2,3</sub>=3.0 Hz, J<sub>3,4</sub>=9.8 Hz, H-3), 4.37-4.08 (m, 5 H, H-5/H-5', H-6", OCH2CHCH2), 3.76 (m, 1 H, H-5/H-5'), 3.75 (dd, 1 H, J<sub>1,2</sub>=1.3 Hz, J<sub>2,3</sub>=3.2 Hz, H-2'), 3.54 (m, 1 H, H-5"), 1.29 (d, 3 H, J<sub>5,6</sub>=6.2 Hz, H-6/ H-6'), 1.16 (s, 3 H, COCH<sub>3</sub>), 1.01 (d, 3 H,  $J_{5.6}$ =6.2 Hz, H-6/H-6'); <sup>13</sup>C NMR (100

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MHz, CDCl<sub>3</sub>): δ 168.9 (COCH<sub>3</sub>), 165.7, 165.6, 165.4, 165.0 164.6, 164.4, 164.3 (7 C, 7 COPh), 117.5 (OCH<sub>2</sub>CHCH<sub>2</sub>), 101.6, 100.5, 96.2 (3 C, C-1, 1', 1"), 77.2, 76.0, 73.0, 72.3, 72.1, 71.6, 71.4, 71.1, 69.7, 69.0, 68.3, 68.2, 67.0, 66.1, 19.2, 17.2, 16.7. Anal. Calcd for C<sub>72</sub>H<sub>66</sub>O<sub>22</sub>: C, 67.38; H, 5.18. Found: C, 67.46; H, 5.03.

2,3,4,6-Tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-benzoyl- $\alpha\text{-L-rhamnopyranosyl-}(1 \rightarrow 3)\text{-}2, 4\text{-di-}\textit{O-benzoyl-}\alpha\text{-L-rhamnopyranosyl} trichloroace$ timidate (14). To a solution 12 (1.2 g, 1 mmol) in 90% acetic acid (10 mL) containing sodium acetate (293 mg, 3 mmol) was added PdCl<sub>2</sub> (89 mg, 0.5 mmol), and the mixture was stirred for 12 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the reaction was complete. The mixture was diluted with dichloromethane (30 mL), washed with water and satd aq sodium bicarbonate. The organic layer was concentrated, and the residue was passed through a short silica gel column with 2:1 petroleum ether-EtOAc as the eluent to give crude 2,3,4,6-tetra-Obenzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -3-O-acetyl-4-O-benzoyl- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 3)$ -2,4-di-O-benzoyl- $\alpha$ , $\beta$ -L-rhamnopyranose (13, 1.05 g, 91.3%). Compound 13 was dissolved in dichloromethane (10 mL), and CCl<sub>3</sub>CN (0.2 mL, 2 mmol) and DBU (27  $\mu$ L, 0.18 mmol) were added. The reaction mixture was stirred for 2 h, at the end of which time TLC (3:1 petroleum ether-ethyl acetate) indicated that the reaction was complete. Concentration of the reaction mixture followed by purification on a silica gel column with 3:1 petroleum ether-EtOAc as the eluent, furnished the trisaccharide donor 14 (1.01 g, 86.0%) as a foamy solid:  $[\alpha]_D + 37.8^\circ$  (c 1.1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.80 (s, 1 H, CNHCCl<sub>3</sub>), 8.19-7.27 (m, 35 H, 7 PhH), 6.47 (d, 1 H, J<sub>1,2</sub>=1.9 Hz, H-1), 5.75 (dd, 1 H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.7 Hz, H-3"), 5.66 (dd, 1 H, J<sub>1,2</sub>=1.9 Hz,  $J_{2,3}=3.3$  Hz, H-2), 5.61 (dd, 1 H,  $J_{3,4}=J_{4,5}=9.7$  Hz, H-4"), 5.51 (dd, 1 H,  $J_{3,4}=J_{4,5}=1.5$ 9.8 Hz, H-4), 5.40 (dd, 1 H, J<sub>1,2</sub>=7.7 Hz, J<sub>2,3</sub>=9.7 Hz, H-2"), 5.20 (d, 1 H, J<sub>1,2</sub>=1.6 Hz, H-1'), 5.18 (dd, 1 H, J<sub>2,3</sub>=3.2 Hz, J<sub>3,4</sub>=9.6 Hz, H-3'), 5.08 (dd, 1 H, J<sub>3,4</sub>=J<sub>4,5</sub>=9.6 Hz, H-4'), 4.60 (d, 1 H, J<sub>1,2</sub>=7.7 Hz, H-1"), 4.54 (dd, 1 H, J<sub>2,3</sub>=3.3 Hz, J<sub>3,4</sub>=9.8 Hz, H-3), 4.30-4.24 (m, 2 H, H-5", H-6"), 4.04-3.91 (m, 2 H, H-5/H-5', H-6"), 3.86 (dd, 1 H, J<sub>1,2</sub>=1.6 Hz, J<sub>2,3</sub>=3.2 Hz, H-2'), 3.61 (m, 1 H, H-5/H-5'), 1.43 (d, 3 H, J<sub>5,6</sub>=6.3 Hz, H-6/H-6'), 1.25 (s, 3 H, COCH<sub>3</sub>), 1.02 (d, 3 H, J<sub>5.6</sub>=6.4 Hz, H-6/H-6').

Anal. Calcd for C<sub>71</sub>H<sub>62</sub>Cl<sub>3</sub>NO<sub>22</sub>: C, 61.45; H, 4.50. Found: C, 61.26; H, 4.41.

Allyl 2,3,4,6-Tetra-*O*-benzoyl-β-D-glucopyranosyl- $(1 \rightarrow 2)$ -3-*O*-acetyl-4-*O*-benzoyl-α-L-rhamnopyranosyl- $(1 \rightarrow 3)$ -2,4-di-*O*-benzoyl-α-L-rhamnopyranosyl- $(1 \rightarrow 2)$ -3,4-di-*O*-benzoyl-α-L-rhamnopyranoside (16). Compound 14 (693 mg, 0.5 mmol) and allyl 3,4-di-*O*-benzoyl-α-L-rhamnopyranoside (15, 206 mg, 0.5 mmol) were dried together under high vacuum for 2 h, then dissolved in anhyd CH<sub>2</sub>Cl<sub>2</sub> (40 mL). TMSOTf (18 µL, 0.1 mmoL) was added dropwise at  $-5^{\circ}$ C with N<sub>2</sub> protection. The reaction mixture was stirred for 3 h, during which time the temperature was gradually raised to ambient temperature. Then the mixture was neutralized with triethylamine, and concentrated. Purification of the residue by column chromatography (2:1 petroleum ether–EtOAc) gave 16 (684 mg, 83.6%) as a syrup:  $[\alpha]_D + 71.2^{\circ}$  (*c* 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.10–7.25 (m, 45 H, 9 PhH), 5.90 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.77 (dd, 1 H, J<sub>2,3</sub>=3.0 Hz, J<sub>3,4</sub>=9.8 Hz, H-3<sub>A</sub>), 5.73 (dd, 1 H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.7 Hz, H-3<sub>D</sub>), 5.67 (dd, 1 H, J<sub>1,2</sub>=1.5 Hz, J<sub>2,3</sub>=3.1 Hz, H-2<sub>B</sub>), 5.60 (dd, 1 H, J<sub>3,4</sub>=J<sub>4,5</sub>=9.8 Hz, H-4<sub>A</sub>), 5.50–5.28 (m, 5 H, H-2<sub>D</sub>, H-4<sub>B</sub>, H-4<sub>D</sub>, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.25 (d, 1 H, J<sub>1,2</sub>=1.6 Hz, H-1<sub>C</sub>), 5.23 (dd, 1

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H,  $J_{2,3}=3.1$  Hz,  $J_{3,4}=9.7$  Hz, H-3<sub>C</sub>), 5.14 (d, 1 H,  $J_{1,2}=1.5$  Hz, H-1<sub>B</sub>), 5.11 (dd, 1 H,  $J_{3,4}=J_{4,5}=9.7$  Hz, H-4<sub>C</sub>), 5.03 (d, 1 H,  $J_{1,2}=1.3$  Hz, H-1<sub>A</sub>), 4.64 (d, 1 H,  $J_{1,2}=7.7$  Hz, H-1<sub>D</sub>), 4.61 (dd, 1 H,  $J_{2,3}=3.1$  Hz,  $J_{3,4}=9.8$  Hz, H-3<sub>B</sub>), 4.31 (dd, 1 H,  $J_{1,2}=1.3$  Hz,  $J_{2,3}=3.0$  Hz, H-2<sub>A</sub>), 4.30–4.07 (m, 6 H), 3.92 (m, 1 H, H-5<sub>C</sub>), 3.90 (dd, 1 H,  $J_{1,2}=1.6$  Hz,  $J_{2,3}=3.1$  Hz, H-2<sub>C</sub>), 3.60 (m, 1 H, H-5<sub>A</sub>), 1.32 (d, 3 H,  $J_{5,6}=6.2$  Hz), 1.28 (s, 3 H, COCH<sub>3</sub>), 1.26 (d, 3 H,  $J_{5,6}=6.4$  Hz), 1.07 (d, 3 H,  $J_{5,6}=6.2$  Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  168.8 (COCH<sub>3</sub>), 165.5, 165.4, 165.3, 165.1, 165.0, 164.9, 164.6, 164.5, 164.3 (9 C, 9 COPh), 117.3 (OCH<sub>2</sub>CHCH<sub>2</sub>), 101.3, 99.8, 99.1, 97.3 (4 C, C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>), 73.8, 73.1, 72.2, 71.7, 71.6, 71.6, 71.5, 71.5, 71.4, 70.7, 69.8, 69.3, 67.9, 67.8, 67.3, 67.0, 66.6, 62.4 (18 C, C-2<sub>A</sub>, 2<sub>B</sub>,2<sub>C</sub>, 2<sub>D</sub>, 3<sub>A</sub>, 3<sub>B</sub>, 3<sub>C</sub>, 3<sub>D</sub>, 4<sub>A</sub>, 4<sub>B</sub>, 4<sub>C</sub>, 4<sub>D</sub>, 5<sub>A</sub>, 5<sub>B</sub>, 5<sub>C</sub>, 5<sub>D</sub>, 6<sub>D</sub>, OCH<sub>2</sub>CHCH<sub>2</sub>), 19.4, 17.2, 17.2, 16.8 (4 C, C-6<sub>A</sub>, 6<sub>B</sub>, 6<sub>C</sub>, COCH<sub>3</sub>).

Anal. Calcd for C<sub>92</sub>H<sub>84</sub>O<sub>28</sub>: C, 67.47; H, 5.17. Found: C, 67.54; H, 5.33.

Allyl 2,3,4,6-Tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -4-O-benzoyl- $\alpha$ -Lrhamnopyranosyl- $(1 \rightarrow 3)$ -2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)$ -3,4-di-Obenzoyl-a-L-rhamnopyranoside (17). To a solution of 16 (654 mg, 0.4 mmol) in anhyd MeOH (50 mL) was added acetyl chloride (1.5 mL) at 0°C. The solution was stoppered in a flask and stirred at room temperature until TLC (3:1 petroleum ether-EtOAc) showed that the starting material disappeared. The solution was neutralized with Et<sub>3</sub>N, then concentrated to dryness. The residue was passed through a short silica gel column to give 17 (603 mg, 94.6%) as a white solid:  $[\alpha]_D + 75.5^\circ$  (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.11–7.32 (m, 45 H, 9 PhH), 5.91 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.79 (dd, 1 H, J<sub>2,3</sub>=3.0 Hz, J<sub>3,4</sub>=9.8 Hz, H-3<sub>A</sub>), 5.68 (dd, 1 H, J<sub>1,2</sub>=0.8 Hz,  $J_{2,3}=3.1$  Hz,  $H-2_B$ ), 5.67 (dd, 1 H,  $J_{2,3}=J_{3,4}=9.6$  Hz,  $H-3_D$ ), 5.62 (dd, 1 H,  $J_{3,4}=1.0$  $J_{4,5}=9.8$  Hz, H-4<sub>A</sub>), 5.53 (dd, 1 H,  $J_{3,4}=J_{4,5}=9.6$  Hz, H-4<sub>D</sub>), 5.48 (dd, 1 H,  $J_{3,4}=J_{4,5}=1.5$ 9.7 Hz, H-4<sub>B</sub>), 5.45 (dd, 1 H, J<sub>1,2</sub>=7.7 Hz, J<sub>2,3</sub>=9.6 Hz, H-2<sub>D</sub>), 5.40-5.26 (m, 2 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.22 (d, 1 H, J<sub>1,2</sub>=0.8 Hz, H-1<sub>B</sub>), 5.13 (d, 1 H, J<sub>1,2</sub>=1.6 Hz, H-1<sub>C</sub>), 5.03 (d, 1 H,  $J_{1,2}$ =1.6 Hz, H-1<sub>A</sub>), 4.80 (dd, 1 H,  $J_{3,4}$ = $J_{4,5}$ =9.9 Hz, H-4<sub>C</sub>), 4.66 (d, 1 H,  $J_{1,2}$ =7.7 Hz, H-1<sub>D</sub>), 4.58 (dd, 1 H,  $J_{2,3}$ =3.1 Hz,  $J_{3,4}$ =9.7 Hz, H-3<sub>B</sub>), 4.32-4.11 (m, 7 H), 3.92 (m, 1 H), 3.78-3.74 (m, 2 H), 3.45 (m, 1 H), 1.33 (d, 3 H, J<sub>5.6</sub>=6.4 Hz), 1.27 (d, 3 H,  $J_{5,6}$ =6.4 Hz), 1.10 (d, 3 H,  $J_{5,6}$ =6.3 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$ 165.9, 165.5, 165.4, 165.3, 165.3, 165.1, 165.0, 164.9, 164.8, 164.5 (9 C, 9 COPh), 117.5 (OCH<sub>2</sub>CHCH<sub>2</sub>), 102.0, 99.5, 99.1, 97.3 (4 C, C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>), 79.3, 73.8, 73.1, 72.1, 71.7, 71.6, 71.6, 71.5, 71.5, 71.4, 70.7, 69.8, 69.2, 68.9, 67.8, 66.9, 66.6, 62.3 (18 C, C-2<sub>A</sub>, 2<sub>B</sub>,2<sub>C</sub>, 2<sub>D</sub>, 3<sub>A</sub>, 3<sub>B</sub>, 3<sub>C</sub>, 3<sub>D</sub>, 4<sub>A</sub>, 4<sub>B</sub>, 4<sub>C</sub>, 4<sub>D</sub>, 5<sub>A</sub>, 5<sub>B</sub>, 5<sub>C</sub>, 5<sub>D</sub>, 6<sub>D</sub>, OCH<sub>2</sub>CHCH<sub>2</sub>), 17.2, 17.1, 16.8 (3 C, C-6<sub>A</sub>, 6<sub>B</sub>, 6<sub>C</sub>).

Anal. Calcd for C<sub>90</sub>H<sub>82</sub>O<sub>27</sub>: C, 67.74; H, 5.18. Found: C, 67.63; H, 5.26.

Allyl 2,3,4-Tri-O-benzoyl- $\beta$ -D-ribofuranosyl- $(1 \rightarrow 3)$ -[2,3,4,6-tetra-O-benzoyl- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ -]4-O-benzoyl- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 3)$ -2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)$ -3,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranoside (19). 2,3,4-Tri-O-benzoyl- $\alpha$ -D-ribofuranosyl trichloroacetimidate (18, 243 mg, 0.4 mmol) and 17 (590 mg, 0.37 mmol) were dried together under high vacuum for 2 h, then dissolved in anhyd CH<sub>2</sub>Cl<sub>2</sub> (40 mL). TMSOTf (18  $\mu$ L, 0.10 mmol) was added dropwise at  $-10^{\circ}$ C with N<sub>2</sub> protection. The reaction mixture was stirred for 3 h, during which time the temperature was gradually warmed to ambient temperature. Then the mixture was neutralized with triethylamine and concentrated to dryness. Purification

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of the residue by column chromatography (1:1 petroleum ether-EtOAc) gave 19 (536 mg, 71.1%) as a foamy solid:  $[\alpha]_{\rm D}$  + 37.1° (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.10–7.19 (m, 60 H, 12 PhH), 6.00 (dd, 1 H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.5 Hz, H-3<sub>D</sub>), 5.94 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.78 (dd, 1 H, J<sub>2,3</sub>=3.2 Hz, J<sub>3,4</sub>=9.8 Hz, H-3<sub>A</sub>), 5.67 (dd, 1 H, J<sub>1,2</sub>=1.7 Hz, J<sub>2.3</sub>=3.3 Hz, H-2<sub>B</sub>), 5.61–5.52 (m, 3 H, H-4<sub>A</sub>, H-4<sub>B</sub>, H-4<sub>D</sub>), 5.47–5.43 (m, 2 H, H-2<sub>D</sub>, H-4<sub>E</sub>), 5.37 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.35 (d, 1 H, J<sub>1,2</sub>=7.7 Hz, H-1<sub>D</sub>), 5.26 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>),  $5.22-5.21(m, 2 H, H-1_C, H-1_E)$ ,  $5.15 (dd, 1 H, J_{3,4}=J_{4,5}=9.8$ Hz, H-4<sub>C</sub>), 5.12 (d, 1 H,  $J_{1,2}$ =1.5 Hz, H-1<sub>B</sub>), 5.00 (d, 1 H,  $J_{1,2}$ =1.5 Hz, H-1<sub>A</sub>), 4.95 (dd, 1 H,  $J_{2,3} = J_{3,4} = 4.0$  Hz, H-3<sub>E</sub>), 4.84 (d, 1 H,  $J_{2,3} = 4.0$  Hz, H-2<sub>E</sub>), 4.61 (dd, 1 H,  $J_{2,3} = 3.4$ Hz, J<sub>3,4</sub>=9.9 Hz, H-3<sub>B</sub>), 4.32–4.06 (m, 10 H), 3.82 (m, 1 H), 3.54–3.39 (m, 2 H), 1.33 (d, 3 H,  $J_{5,6}=6.3$  Hz), 1.29 (d, 3 H,  $J_{5,6}=6.4$  Hz), 0.75 (d, 3 H,  $J_{5.6}=6.3$  Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 165.6, 165.4, 165.3, 165.1, 165.0, 164.9, 164.9, 164.9, 164.8, 164.5, 164.3, 163.9 (12 C, 12 COPh), 117.5 (OCH<sub>2</sub>CHCH<sub>2</sub>), 100.0 (1 C, C-1<sub>A/B/C</sub>,  $J_{C1-H1} = 173 \text{ Hz}$ ), 99.5 (1 C, C-1<sub>E</sub>,  $J_{C1-H1} = 162 \text{ Hz}$ ), 99.1 (2 C, C-1<sub>A/B/C</sub>,  $J_{C1-H1} = 171 \text{ Hz}$ ) Hz), 97.2 (1 C, C-1<sub>D</sub>,  $J_{C1-H1}$  = 164 Hz), 17.2, 17.2, 16.5 (3 C, C-6<sub>A</sub>, 6<sub>B</sub>, 6<sub>C</sub>). Anal. Calcd for C<sub>116</sub>H<sub>102</sub>O<sub>34</sub>: C, 68.29; H, 5.09. Found: C, 68.41; H, 5.03.

Allyl  $\beta$ -D-Ribofuranosyl- $(1 \rightarrow 3)$ - $[\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ ]- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 3)$ - $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)$ - $\alpha$ -L-rhamnopyranoside (20). Pentasaccharide 19 (500 mg, 0.25 mmol) was dissolved in a saturated solution of ammonia in MeOH (10 mL). After 4 days at rt, the reaction mixture was concentrated and the residue was purified by chromatography on Sephadex LH-20 (MeOH) to afford 20 as a foamy solid (151 mg, 77.8%);  $[\alpha]_{\rm D} - 46.1^{\circ}$  (c 1.0, D<sub>2</sub>O); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$ 5.95 (m, 1 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.43 (s, 1 H, H-1), 5.41–5.30 (m, 2 H, OCH<sub>2</sub>CHCH<sub>2</sub>), 5.05 (d, 1 H,  $J_{1,2}$  5.1 Hz, H-1), 4.94 (s, 2 H, H-1), 4.62 (d, 1 H,  $J_{1,2}$  10.3 Hz, H-1); <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O): δ 105.8, 104.8, 104.4, 102.9, 101.9 (5C, C-1); MS (m/z) Calcd for C<sub>32</sub>H<sub>54</sub>O<sub>22</sub>: 790.31 [M]<sup>+</sup>. Found: 813.52[M+Na]<sup>+</sup>.

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#### REFERENCES

- 1. Previato, J.O.; Jones, C.; Stephan, M.P.; Almeida, L.P.A.; Mendonca-Previato, L. Structures of the repeating oligosaccharide from the lipopolysaccharide of the nitrogen-fixing bacterium Acetobacter diazotrophicus strain PAL 5. Carbohydr. Res. 1997, 298, 311-318.
- 2. Lima, E.; Boddey, R.M.; Döbereiner, J. Quantification of biological nitrogen fixation associated with sugar cane using a nitrogen-15 aided nitrogen balance. Soil Biol. Biochem. 1987, 19, 165-170.
- Battisti, L.; Lara, J.C.; Leigh, J.A. Specific succinoglycan plays an important role in 3. root nodulation development. Proc. Natl. Acad. Sci. U. S. A. 1992, 89, 5625-5629.

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- Marino-Albernas, J.; Harris, S.L.; Varma, V.; Pinto, B.M. Convergent synthesis of an elusive hexasaccharide corresponding to the cell-wall polysaccharide of the β-hemolytic *Streptococcus* Group A. Carbohydr. Res. **1993**, *298*, 245–257.
- Wang, W.; Kong, F. Highly regio- and steroselective synthesis of mannose-containing oligosaccharides with acetobromo sugars as the donors and partially protected mannose derivatives as the acceptors via sugar orthoester intermediates. Angew. Chem., Int. Ed. 1999, 38, 1247–1250.
- Wang, W.; Kong, F. New synthetic methodology for regio-and stereoselective synthesis of oligosaccharides via sugar orthoester intermediates. J. Org. Chem. 1998, 63, 5744–5755.
- Du, Y.; Kong, F. Regio- and stereoselective synthesis of rhamnose-containing oligosaccharides via sugar-sugar orthoester formation and rearrangement. J. Carbohydr. Chem. 1999, 18, 655–664.
- 8. Zhu, Y.; Kong, F. Synthesis of  $\alpha$ -(1 $\rightarrow$ 6)-linked mannose dodecasaccharide and  $\beta$ -(1 $\rightarrow$ 6)-linked glucooctaose using sugar trichloroacetimidates as the donor and unprotected and partially protected glycosides as the acceptors. Synlett **2000**, 663–669.
- Chiu-Machado, I.; Castro-Palomino, J.C.; Madrazo-Aloso, O.; Loipeteguipalucious, C.; Verez-Bencomo, V. Synthesis of ribofuranosides by catalysis with Lewis acids. Glycosidation versus transacetylation. J. Carbohydr. Chem. **1995**, *14*, 551–561.
- 10. Schmidt, R.R.; Kinzy, W. Anomeric-oxygen activation for glycoside synthesis: The trichloroacetimidate method. Adv. Carbohydr. Chem. Biochem. **1994**, *50*, 21–125.

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